

INFEZIONI DA MDR

Il ruolo del Laboratorio

LE MALATTIE INFETTIVE OGGI

San Fermo della Battaglia, 12 Settembre 2019

Auditorium Ospedale Sant'Anna – ASST Lariana



1- RAPIDITA' NEL FORNIRE UN REFERTO

2- FORNIRE UN REFERTO AGGIORNATO

3- SORVEGLIANZA ED
EPIDEMIOLOGIA LOCALE



1- RAPIDITA' NEL FORNIRE UN REFERTO

2- FORNIRE UN REFERTO AGGIORNATO

3- SORVEGLIANZA



1 – IDENTIFICAZIONE BATTERICA
CON SPETTROMETRIA DI MASSA

2 – TEST RAPIDI BIOLOGIA MOLECOLARE
PER RESISTENZE

3 – TEST RAPIDI IMMUNOCROMATOGRAPHICI
PER RESISTENZE

Diagnostica microbiologica della sepsi

Metodi tradizionali

- Giorno 1: prelievi ed incubazione dei flaconi.
- Giorno 2: positivizzazione dei flaconi
- Giorno 3: crescita delle colonie
- Giorno 4: identificazione batterica (biochimica) ed antibiogramma

Spettrometria di massa MALDI - TOF

- Giorno 1: prelievi ed incubazione dei flaconi
- Giorno 2: positivizzazione dei flaconi con identificazione batterica (spettrometria di massa) da flacone positivo.
- Giorno 3: antibiogramma.

Identificazione diretta da emocoltura positiva mediante spettrometria di massa (MALDI-TOF)



Position paper AMCLI sulla possibilità di *Identificazione diretta dei microrganismi da emocoltura positiva con il dispositivo Vitek[®] MS*

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Position paper AMCLI sulla possibilità di *Identificazione diretta dei microrganismi da emocoltura positiva con il dispositivo Vitek[®] MS*

- L'utilizzo della tecnologia MALDI-TOF MS per l'identificazione diretta dei batteri aerobi da emocoltura positiva si è confermata essere una possibilità diagnostica di grande interesse, essendo in grado di fornire (soprattutto per le specie microbiche maggiormente critiche sul piano clinico-terapeutico) una importante anticipazione nell'identificazione batterica con livelli di accuratezza adeguati.
- Fra i protocolli operativi considerati nella nostra valutazione si sono in particolare dimostrati altamente performanti la Centrifugazione e Semina con incubazione del terreno solido per 4 ore e la **Semina Diretta con incubazione protratta per 5 ore.**

RESEARCH ARTICLE

Open Access



Identification by mass spectrometry and automated susceptibility testing from positive bottles: a simple, rapid, and standardized approach to reduce the turnaround time in the management of blood cultures

Carola Mauri¹, Luigi Principe¹, Silvia Bracco^{1,2}, Elisa Meroni¹, Nicoletta Corbo¹, Beatrice Pini¹ and Francesco Luzzaro^{1*} 

Abstract

Background

Speeding up identification and antimicrobial susceptibility testing (AST) is of foremost importance in the management of blood cultures. Here, we describe a simple, rapid, and standardized approach based on a very short-term incubation on solid medium from positive blood cultures followed by MALDI-TOF mass spectrometry identification and automated AST. The aim of the study was to evaluate the impact in the laboratory practice of this new procedure with respect to that previously used (standard method) by comparing TAT and cumulative percentage of final reports to clinicians.

BMC Infectious Diseases

Results

Compared with the standard method, the new procedure gave correct organism identification at genus or species level in 98.4% of monomicrobial samples. AST resulted in 97.7% essential agreement and 98.1% categorical agreement, with 0.9% minor errors, 1.0% major error, and 1.5% very major errors. The mean turnaround time to identification and AST was 61.4 h by using the new method compared to 83.1 h by using standard procedure. Concerning cumulative percentages of final reports, approximately a third of results were available at 48 h from the check-in of the sample when using the new procedure, whereas no final reports were ready at the same time with the standard method.

Conclusions

The new procedure, based on rapid identification by MALDI-TOF MS and automated susceptibility testing performed following short-term incubation cultures on solid agar plates inoculated from positive blood cultures, represents a simple, standardized, and workflow-friendly approach that allows faster and reliable results essentially without additional hands-on processing time and costs. The most relevant conclusion from our experience is that the new procedure permits at least a one day reduction in TAT in the management of bloodstream infections, especially when the most frequently isolated pathogens are involved. The introduction of this method in the microbiology laboratory might facilitate an appropriate species-specific therapy with potential downstream impact on the development of resistance and improved patient outcomes.

Xpert® MRSA/SA BC

Rilevamento accurato dell'MRSA e dello SA in campioni di emocolture positive in circa un'ora



Il saggio Xpert MRSA/SA per emocoltura include reagenti per il rilevamento dell'MRSA e dell'SA e un controllo del trattamento del campione (SPC) per verificare l'efficacia del trattamento dei batteri bersaglio e per monitorare la presenza di inibitori nella reazione PCR. I test di controllo della sonda (PCC) verificano la reidratazione dei reagenti, il riempimento della provetta PCR nella cartuccia, l'integrità della sonda e la stabilità del colorante.

I primer e le sonde nel saggio Xpert MRSA/SA per emocoltura rilevano sequenze brevettate per la proteina A stafilococcica (*spa*), il gene per la resistenza alla meticillina/oxacillina (*mecA*) e il cromosoma nella cassetta stafilococcica *mec* (*SCCmec*) inserito nel sito *attB* cromosomico dell'SA.

Comparison of the Next-Generation Xpert MRSA/SA BC Assay and the GeneOhm StaphSR Assay to Routine Culture for Identification of *Staphylococcus aureus* and Methicillin-Resistant *S. aureus* in Positive-Blood-Culture Broths

Blake W. Buchan,^{a,b} Stephen Allen,^c  Carey-Ann D. Burnham,^d Erin McElvania TeKippe,^d Thomas Davis,^e Michael Levi,^f Donna Mayne,^g Preeti Pancholi,^h Ryan F. Relich,^c Richard Thomson,ⁱ Nathan A. Ledebøer^{a,b}

Medical College of Wisconsin, Milwaukee, Wisconsin, USA^a; Dynacare Laboratories, Milwaukee, Wisconsin, USA^b; Indiana University School of Medicine, Indianapolis, Indiana, USA^c; Washington University School of Medicine, St. Louis, Missouri, USA^d; Wishard Memorial Hospital, Indianapolis, Indiana, USA^e; Montefiore Medical Center, Bronx, New York, New York, USA^f; Sacred Heart Hospital, Pensacola, Florida, USA^g; The Ohio State University Wexner Medical Center, Columbus, Ohio, USA^h; NorthShore University HealthSystem, Evanston, Illinois, USAⁱ

Comparison of the Next-Generation Xpert MRSA/SA BC Assay and the GeneOhm StaphSR Assay to Routine Culture for Identification of *Staphylococcus aureus* and Methicillin-Resistant *S. aureus* in Positive-Blood-Culture Broths

A bloodstream infection with *Staphylococcus aureus*, including methicillin-resistant *S. aureus* (MRSA), is a serious condition that carries a high mortality rate and is also associated with significant hospital costs. The rapid and accurate identification and differentiation of methicillin-susceptible *S. aureus* (MSSA) and MRSA directly from positive blood cultures has demonstrated benefits in both patient outcome and cost-of-care metrics. We compare the next-generation Xpert MRSA/SA BC (Xpert) assay to the GeneOhm StaphSR (GeneOhm) assay for the identification and detection of *S. aureus* and methicillin resistance in prospectively collected blood culture broths containing Gram-positive cocci. All results were compared to routine bacterial culture as the gold standard. Across 8 collection and test sites, the Xpert assay demonstrated a sensitivity of 99.6% (range, 96.4% to 100%) and a specificity of 99.5% (range, 98.0% to 100%) for identifying *S. aureus*, as well as a sensitivity of 98.1% (range, 87.5% to 100%) and a specificity of 99.6% (range, 98.3% to 100%) for identifying MRSA. In comparison, the GeneOhm assay demonstrated a sensitivity of 99.2% (range, 95.2% to 100%) and a specificity of 96.5% (range, 89.2% to 100%) for identifying *S. aureus*, as well as a sensitivity of 94.3% (range, 87.5% to 100%) and a specificity of 97.8% (range, 96.1% to 100%) for identifying MRSA. Five of six cultures falsely reported as negative for MRSA by the GeneOhm assay were correctly identified as positive by the Xpert assay, while one culture falsely reported as negative for MRSA by the Xpert assay was correctly reported as positive by the GeneOhm assay.



Contents lists available at ScienceDirect

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journal homepage: www.elsevier.com/locate/diagmicrobio



Clinical Studies

Influence of GeneXpert MRSA/SA test implementation on clinical outcomes of *Staphylococcus aureus* bacteremia — a before–after retrospective study



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ABSTRACT

Use of GeneXpert MRSA/SA in diagnostic algorithms of *Staphylococcus aureus* bacteremia may influence both patients' clinical outcomes and antibiotic stewardship. We evaluated these outcomes in a retrospective cohort before (1/6/2015–31/5/2016) and after (1/6/2016–31/8/2017) the introduction of the test in adult patients with Gram-positive cocci in clusters in blood cultures. We included 254 patients (125 preintervention, 129 postintervention). No significant difference in 30-day mortality or clinical success was demonstrated between periods. Appropriate antibiotic therapy rates were significantly higher in the postintervention group, and vancomycin use was significantly reduced (80.6% vs 53.6%, $P < 0.01$; 2.3 ± 0.38 vs 2.98 ± 1.02 defined daily doses/100 patient days, $P = 0.026$, respectively). Appropriate beta-lactam use was also significantly higher (56.7% postintervention vs 23.1% preintervention, $P < 0.01$). Use of GeneXpert MRSA/SA test has a positive effect on antibiotic stewardship measures, though it has no significant effect on clinical outcomes including mortality in this fatal infection.

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Cepheid - GeneXpert Carba-R Assay



Risultato in circa 1 ora

*bla*_{OXA-48}

OXA-48

*bla*_{IMP-1}

OXA-162

IMP-1 subgroup

OXA-163

IMP-1

IMP-6

OXA-181

IMP-3

IMP-25

OXA-204

IMP-10

IMP-30

*bla*_{KPC}

KPC

*bla*_{VIM}

VIM

*bla*_{NDM-1}

NDM

NG-Test CARBA 5

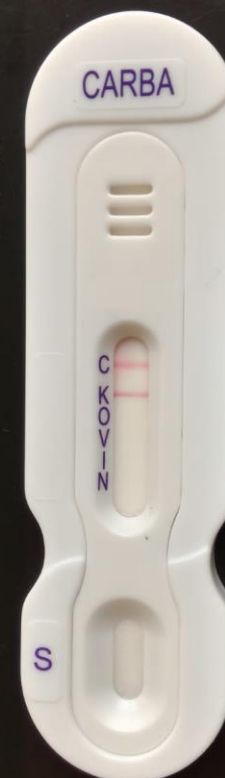
Solo per uso diagnostico in vitro

Test rapido per la ricerca delle carbapenemasi KPC, OXA, VIM, IMP e NDM nelle colonie batteriche da coltura

Solo per uso professionale



NG-Test CARBA 5 è un test immunocromatografico rapido, con lettura visiva, per la rilevazione delle 5 carbapenemasi KPC, OXA, VIM, IMP e NDM in una singola colonia batterica ottenuta da coltura.



05/set/2019

Simplified Testing Method for Direct Detection of Carbapenemase-Producing Organisms from Positive Blood Cultures Using the NG-Test Carba 5 Assay.

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ABSTRACT

We directly tested 484 organisms from clinical samples (n=310) and simulated (n=174) positive blood cultures using NG-Test Carba 5 assay for carbapenemase-producing *Enterobacterales* detection. The assay identified all but 4 of the KPC (170/171), OXA-48-like (22/22), VIM (19/21), and NDM (14/15) producers with no false positives. Among the clinical *Klebsiella pneumoniae* organisms tested, 122 of 123 KPC, 1 of 1 OXA-48-like, and 1 of 2mVIM producers were detected by the assay. Some VIM and NDM producers yielded scant but still-readable bands with the assay. No organisms produced the IMPs that the assay was designed to detect.



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EUCAST

EUROPEAN COMMITTEE
ON ANTIMICROBIAL
SUSCEPTIBILITY TESTING

European Society of Clinical Microbiology and Infectious Diseases

<http://www.eucast.org/>

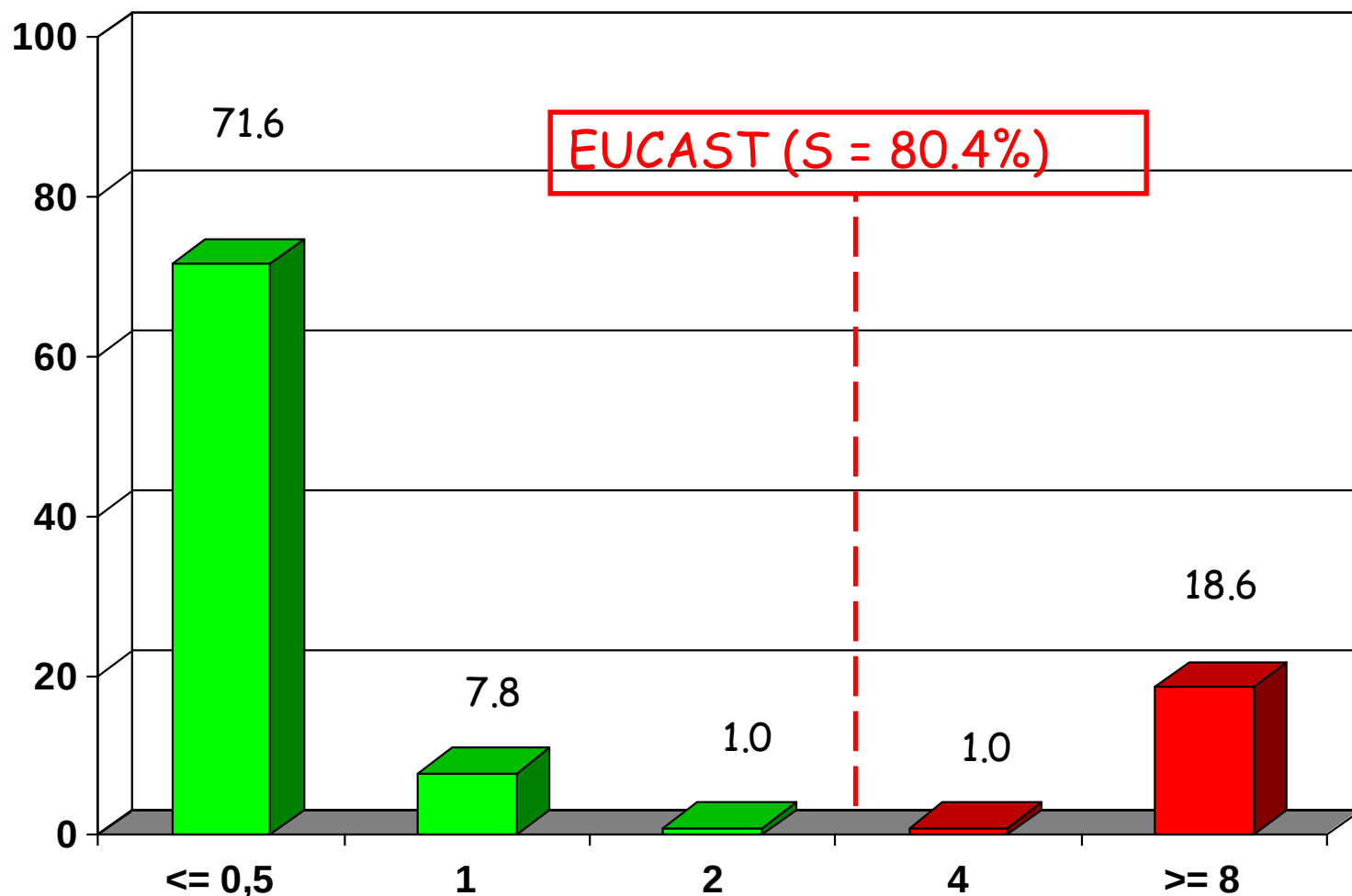
VECCHI ANTIBIOTICI

- COLISTINA
- FOSFOMICINA

NUOVI ANTIBIOTICI

- CEFTAZIDIME-
AVIBACTAM
- CEFTOLOZANO-
TAZOBACTAM

K. pneumoniae KPC + (n=102): sensibilità alla colistina



Colistin resistance superimposed to endemic carbapenem-resistant *Klebsiella pneumoniae*: a rapidly evolving problem in Italy, November 2013 to April 2014

M. Monaco^{1,2}, T Giani^{2,3}, M Raffone^{1,4}, F Arena³, A Garcia-Fernandez¹, S Pollini³, Network EuSCAPE-Italy⁵, H Grundmann⁶, A Pantosti (annalisa.pantosti@iss.it)¹, G M Rossolini^{3,7,8}
www.eurosurveillance.org published on 23 October 2014

Outbreak of Colistin-Resistant, Carbapenemase-Producing *Klebsiella pneumoniae*: Are We at the End of the Road?

David van Duin,^a Yohei Doi^b

Journal of Clinical Microbiology

October 2015 Volume 53 Number 10

Large Nosocomial Outbreak of Colistin-Resistant, Carbapenemase-Producing *Klebsiella pneumoniae* Traced to Clonal Expansion of an *mgrB* Deletion Mutant

Tommaso Giani,^a Fabio Arena,^a Guendalina Vaggelli,^b Viola Conte,^a Adriana Chiarelli,^a Lucia Henrici De Angelis,^a Rossella Fornaini,^c Maddalena Grazzini,^d Fabrizio Niccolini,^d Patrizia Pecile,^b Gian Maria Rossolini^{a,b,e,f}

Antimicrobial susceptibility testing of colistin – evaluation of seven commercial MIC products against standard broth microdilution for *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Acinetobacter* spp.

E. Matuschek*, J. Åhman, C. Webster, G. Kahlmeter

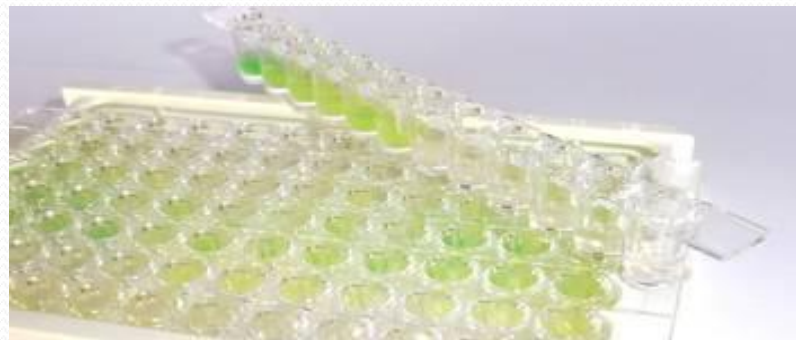
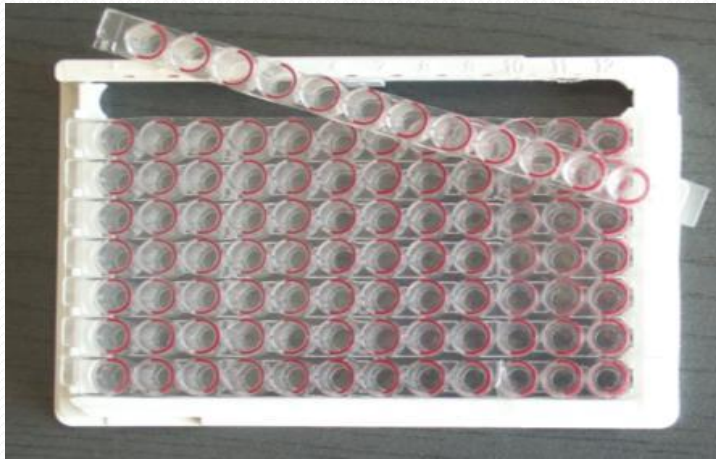
EUCAST Development Laboratory, Växjö, Sweden

Results: The colistin BMD products correlated well with reference tests, in particular for Sensititre and the two MICRONAUT products (essential agreement $\geq 96\%$: 66/69 (96%, CI 88–99%), 72/75 (96%, CI 88–99%) and 74/75 (99%, CI 92–100%)). The results were somewhat poorer for the BMD products SensiTest and UMIC: EA 88% (51/58, CI 77–95%) and 82% (61/74, CI 72–89%), respectively), and considerably poorer for the gradient tests (EA 43–71% depending on gradient test and Mueller-Hinton agar manufacturer). The gradient tests generally underestimated colistin MICs, resulting in a significant number of false susceptible results (9–18 of total 75 tests, compared with 1–3 for the BMD products).

I test di microdiluizione in brodo correlano bene con i test di riferimento, in particolare nel caso di Sensititre e Micronaut.

Risultati considerevolmente più bassi sono stati ottenuti nel caso dei test di diffusione con gradiente (essential agreement tra il 43 e il 71%). Questi test generalmente sottostimano la MIC per la colistina, determinando un significativo numero di risultati falsamente sensibili.

Sistemi di microdiluzione in brodo per singolo antibiotico



1	2	3	4	5	6	7	8	9	10	11	12
GC	COL 0.0625	COL 0.125	COL 0.25	COL 0.5	COL 1	COL 2	COL 4	COL 8	COL 16	COL 32	COL 64

Discrepancies in fosfomycin susceptibility testing of KPC-producing *Klebsiella pneumoniae* with various commercial methods

Giulio Camarlinghi ^a, Eva Maria Parisio ^a, Alberto Antonelli ^b, Maria Nardone ^a, Marco Coppi ^b, Tommaso Giani ^b, Romano Mattei ^a, Gian Maria Rossolini ^{b,c,*}

Fosfomycin susceptibility testing with Sensititre, Vitek2, Etest, Mic Strip Test and disk diffusion methodologies was compared versus reference agar dilution method (AD) with 78 clinical isolates of KPC-producing *Klebsiella pneumoniae*. All methodologies showed a Categorical Agreement and Essential Agreement of $\leq 69\%$ and $\leq 72\%$, respectively, revealing a very low concordance with AD.

Tutte le metodologie testate (Sensititre, Vitek 2, Etest, MIC Strip Test, diffusione da disco) mostravano una bassa concordanza rispetto al metodo di riferimento (agar- diluizione).

In particolare, Sensititre ed Etest avevano alte percentuali di VME e presumibilmente tendono a sottostimare i valori.

Vitek 2, il MIC Strip Test e la diffusione da disco avevano alte percentuali di ME e quindi tendono a sovrastimare i valori di MIC.

Diagnostic Microbiology and Infectious Disease xxx (2018) xxx-xxx



AD Fosfomicin 0.25-256

Device for fosfomicin susceptibility testing with the agar dilution method.

CONFIGURATION

Fosfomicin MIC range: 0.25 - 256 µg/mL

A	256	128	64	32
B	16	8	4	2
C	1	0.5	0.25	Control

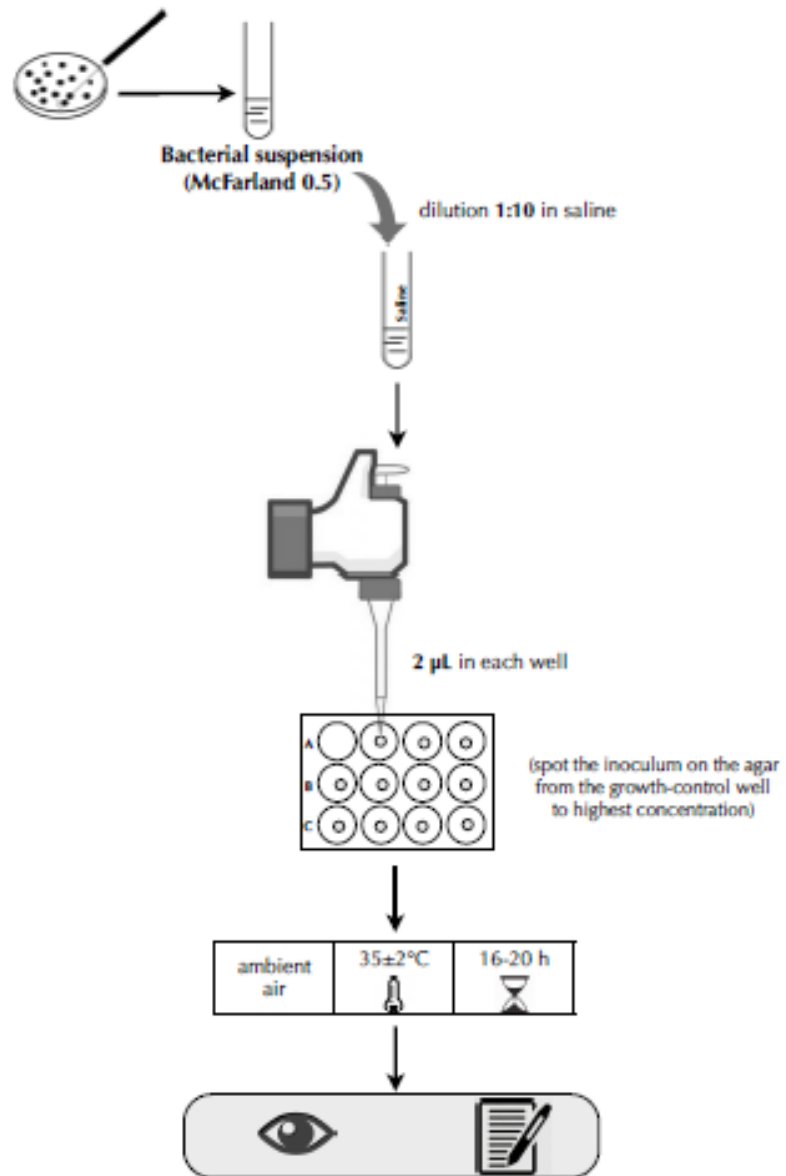
Growth-control: No antimicrobial agent in the well.

PRINCIPLE OF THE METHOD

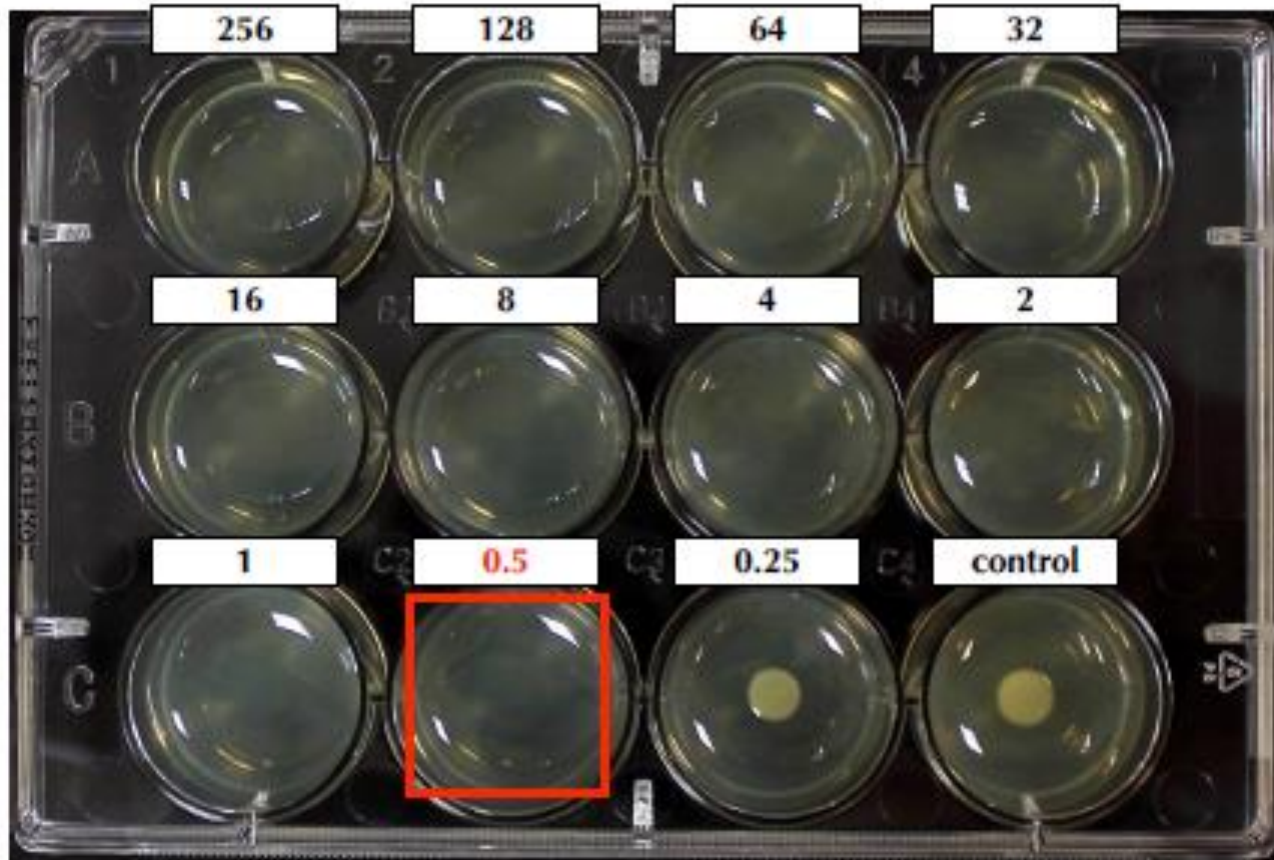
All wells are inoculated with a standardized microbial suspension. After incubation the MIC result is read and interpreted.



AD Fosfomycin Workflow

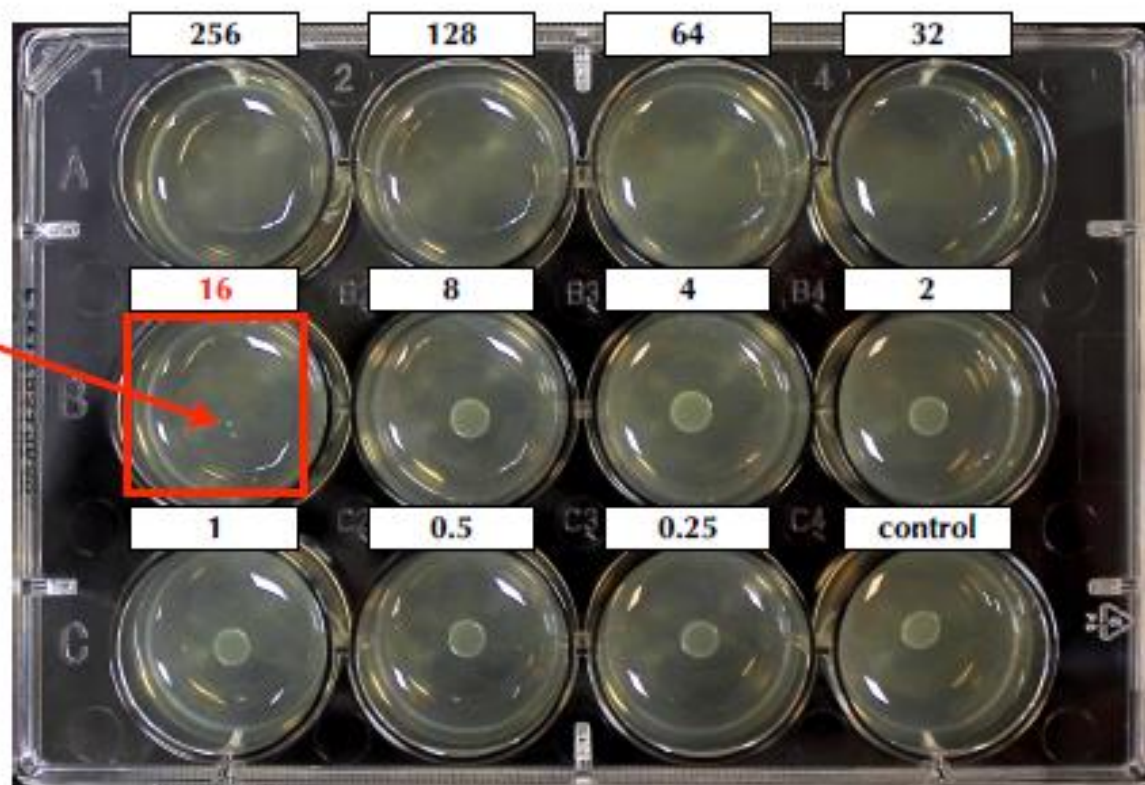


Fosfomycin concentration ($\mu\text{g/mL}$)

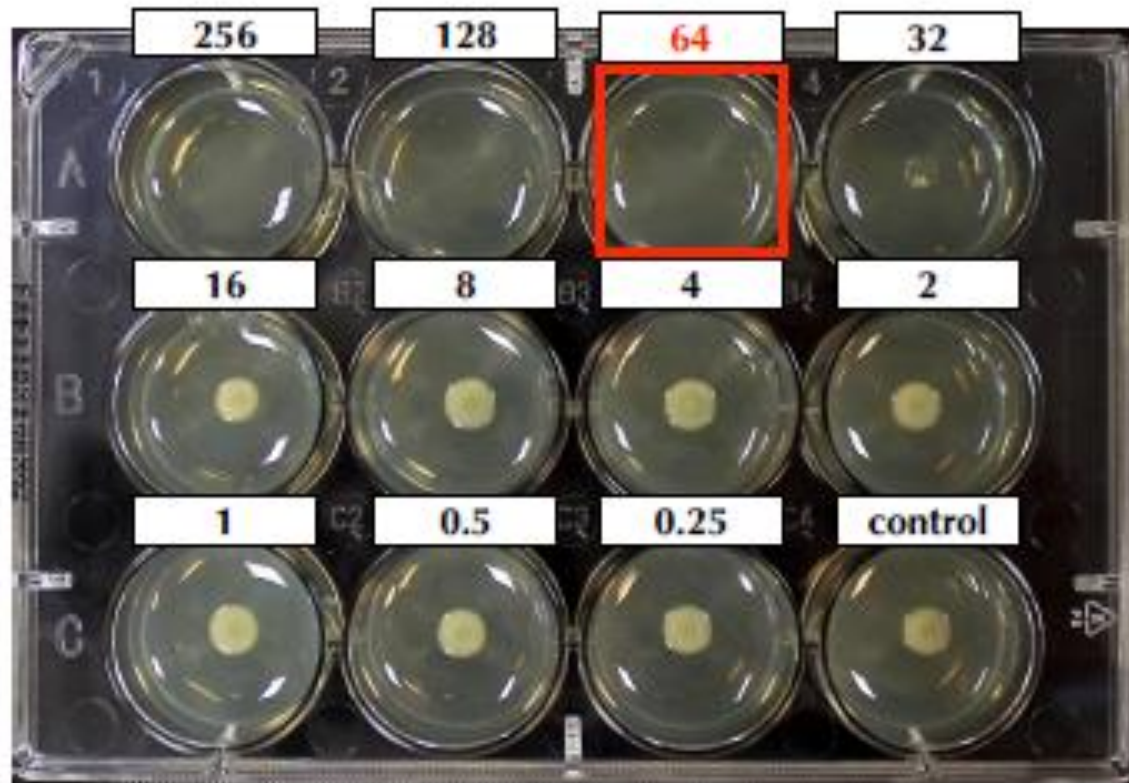


MIC = 0.5 $\mu\text{g/mL}$

Ignore single colonies



MIC = 16 $\mu\text{g/mL}$



MIC = 64 $\mu\text{g/mL}$

Enterobacterales*

Expert Rules and Intrinsic Resistance Tables

Miscellaneous agents	MIC breakpoints (mg/L)			Disk content (µg)	Zone diameter breakpoints (mm)		
	S ≤	R >	ATU		S ≥	R <	ATU
Chloramphenicol	8	8		30	17	17	
Colistin ¹	2	2			Note ^A	Note ^A	
Daptomycin	-	-			-	-	
Fosfomycin iv	32 ²	32 ²		200 ^B	24 ^{C,D}	24 ^{C,D}	
Fosfomycin oral (uncomplicated UTI only)	32 ²	32 ²		200 ^B	24 ^{C,D}	24 ^{C,D}	
Fusidic acid	-	-			-	-	
Metronidazole	-	-			-	-	
Nitrofurantoin (uncomplicated UTI only), <i>E. coli</i>	64	64		100	11	11	
Nitroxoline (uncomplicated UTI only), <i>E. coli</i>	16	16		30	15	15	
Rifampicin	-	-			-	-	
Spectinomycin	-	-			-	-	
Trimethoprim (uncomplicated UTI only)	2	4		5	18	15	
Trimethoprim-sulfamethoxazole ³	2	4		1.25-23.75	14	11	

Enterobacterales*

Expert Rules and Intrinsic Resistance Tables

Miscellaneous agents

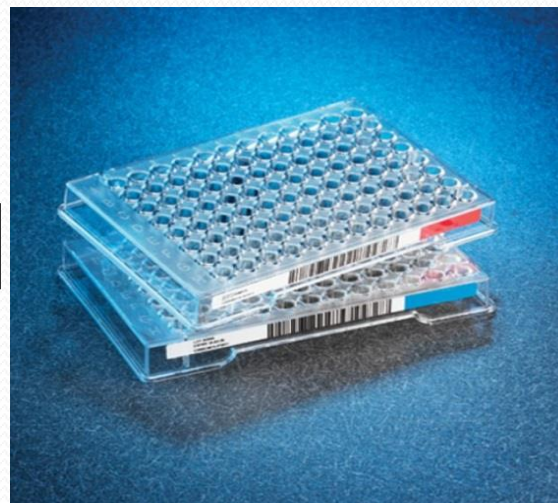
EUCAST Clinical Breakpoint Tables v. 9.0, valid from 2019-01-01

Cephalosporins ¹	MIC breakpoints (mg/L)			Disk content (µg)	Zone diameter breakpoints (mm)		
	S ≤	R >	ATU		S ≥	R <	ATU
Cefaclor	-	-			-	-	
Cefadroxil (uncomplicated UTI only)	16	16		30	12	12	
Cefalexin (uncomplicated UTI only)	16	16		30	14	14	
Cefazolin	-	-			-	-	
Cefepime	1	4		30	27	24	
Cefixime (uncomplicated UTI only)	1	1		5	17	17	
Cefotaxime	1	2		5	20	17	
Cefoxitin (screen) ²	NA	NA		30	19	19	
Cefpodoxime (uncomplicated UTI only)	1	1		10	21	21	
Ceftaroline	0.5	0.5		5	23	23	22-23
Ceftazidime	1	4		10	22	19	
Ceftazidime-avibactam	8 ³	8 ³		10-4	13	13	
Ceftibuten (UTI only)	1	1		30	23	23	
Ceftobiprole	0.25	0.25		5	23	23	
Ceftolozane-tazobactam	1 ⁴	1 ⁴		30-10	23	23	
Ceftriaxone	1	2		30	25	22	

NUOVI ANTIBIOTICI E SPETTRO D'AZIONE

Antibiotico	ESBL	CRE	MDR Pseudomonas	MDR Acinetobacter
Cefiderocol	SI	KPC e NDM	SI	SI
Ceftolozano/Tazobactam	SI	NO	SI	NO
Ceftazidime/Avibactam	SI	KPC e OXA-48 (NO MBL)	SI	NO
Ceftarolina/Avibactam	SI	KPC e OXA-48 (NO MBL)	NO	NO
Aztreonam/Avibactam	SI	MBL	SI	NO
Meropenem/Vaborbactam	SI	KPC	NO	NO
Imipenem/Relebactam	SI	KPC e OXA-48 (NO MBL)	NO	NO
Plazomicina	SI	KPC (NO NDM)	NO	NO
Everacyclina	SI	KPC	NO	SI

Microdiluizione in brodo



Pannello Gram-negativi MDR

	1	2	3	4	5	6	7	8	9	10	11	12
A	MERO 0.12	MERO 0.25	MERO 0.5	MERO 1	MERO 2	MERO 4	MERO 8	MERO 16	AMI 4	AMI 8	AMI 16	AMI 32
B	GEN 0.5	GEN 1	GEN 2	GEN 4	GEN 8	AZT 0.5	AZT 1	AZT 2	AZT 4	AZT 8	AZT 16	AZT 32
C	CIP 0.06	CIP 0.12	CIP 0.25	CIP 0.5	CIP 1	CIP 2	P/T4 1/4	P/T4 2/4	P/T4 4/4	P/T4 8/4	P/T4 16/4	P/T4 32/4
D	AUGC 4/2	AUGC 8/2	AUGC 16/2	AUGC 32/2	AUGC 64/2	C/T 0.5/4	C/T 1/4	C/T 2/4	C/T 4/4	C/T 8/4	C/T 16/4	C/T 32/4
E	COL 0.25	COL 0.5	COL 1	COL 2	COL 4	COL 8	FOT 0.5	FOT 1	FOT 2	FOT 4	FOT 8	TOB 1
F	TGC 0.25	TGC 0.5	TGC 1	TGC 2	TGC 4	SXT 1/19	SXT 2/38	SXT 4/76	SXT 8/152	TOB 2	TOB 4	TOB 8
G	TAZ 0.5	TAZ 1	TAZ 2	TAZ 4	TAZ 8	TAZ 16	CZA 0.5/4	CZA 1/4	CZA 2/4	CZA 8/4	CZA 16/4	NEG
H	IMI 0.5	IMI 1	IMI 2	IMI 4	IMI 8	IMI 16	ETP 0.12	ETP 0.25	ETP 0.5	ETP 1	ETP 2	POS



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ORIGINAL ARTICLE

Potential Role of Active Surveillance in the Control of a Hospital-Wide Outbreak of Carbapenem-Resistant *Klebsiella pneumoniae* Infection

Debby Ben-David, MD; Yasmin Maor, MD; Nathan Keller, MD; Gili Regev-Yochay, MD; Ilana Tal, MS; Dalit Shachar, RN; Amir Zlotkin, PhD; Gill Smollan, MD; Galia Rahav, MD

The optimal timing of active surveillance and the interval between the performance of one culture and the next are not well defined.

Most institutions obtain admission culture samples as well as weekly culture samples.



Mario Sarti

SERVIZIO SANITARIO REGIONALE
EMILIA-ROMAGNA
Azienda Unità Sanitaria Locale di Modena



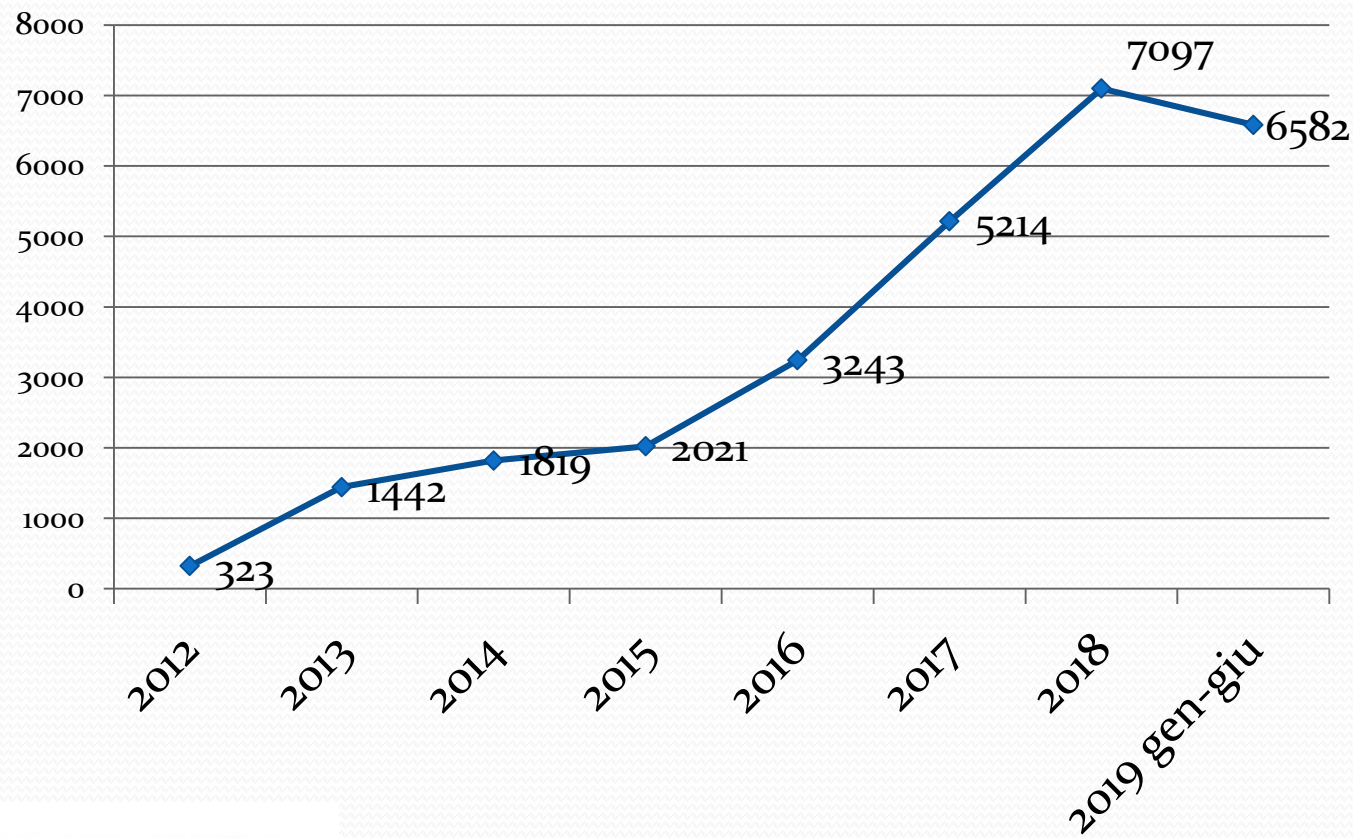
INFECTION CONTROL AND HOSPITAL EPIDEMIOLOGY APRIL 2014, VOL. 35, NO. 4

ORIGINAL ARTICLE

Clinical Microbiology Costs for Methods of Active Surveillance for *Klebsiella pneumoniae* Carbapenemase–Producing Enterobacteriaceae

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Costi D. Sifri, MD;¹ Kevin C. Hazen, PhD^{2,a}

Tampone rettale screening CRE



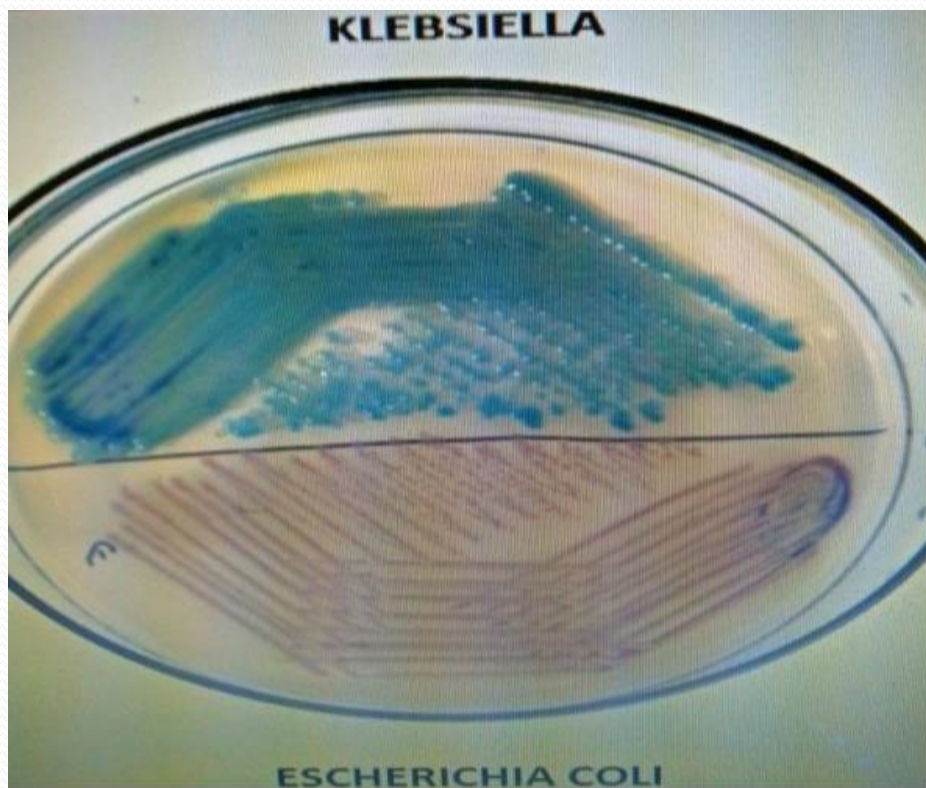
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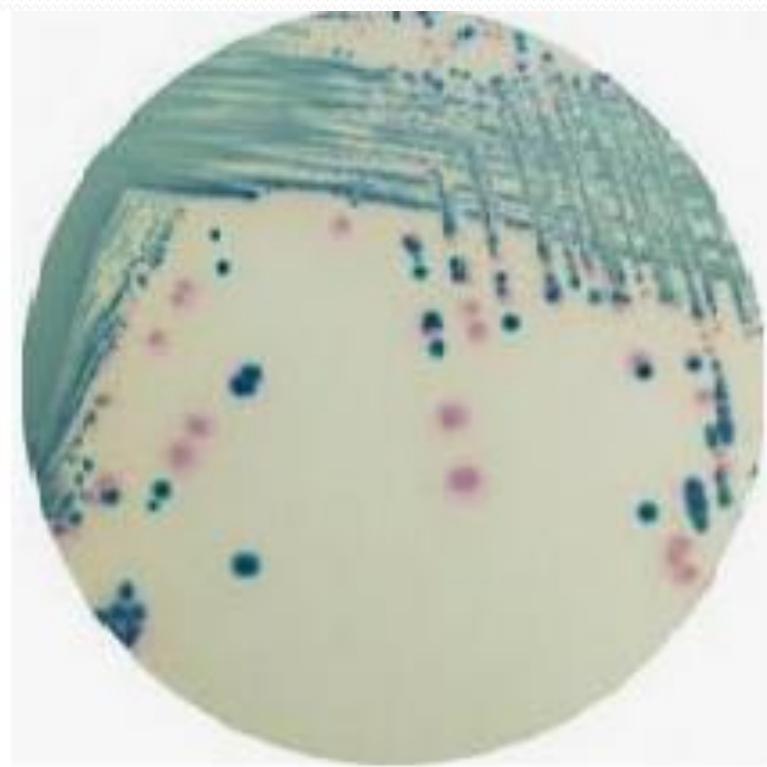
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KLEBSIELLA



ESCHERICHIA COLI

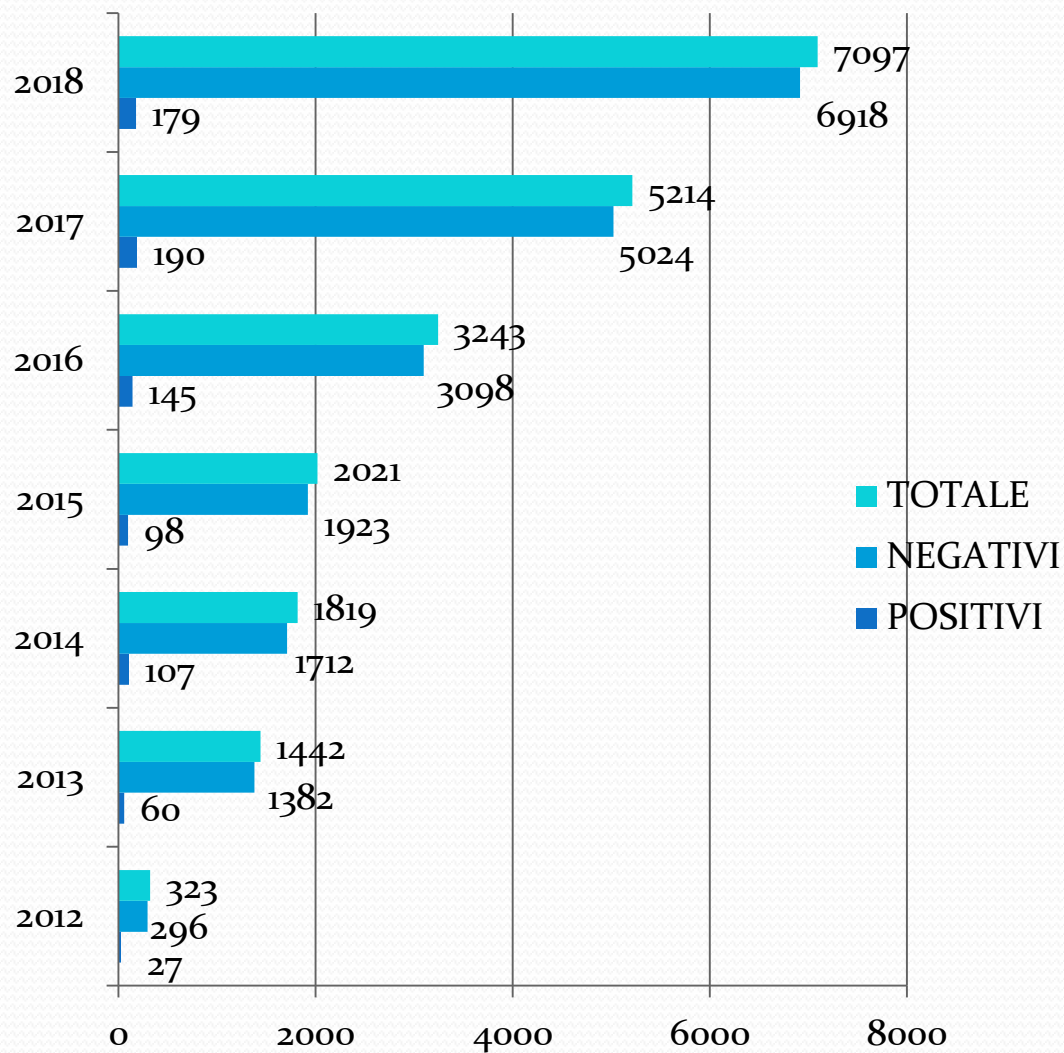


WASP

Walk away specimen processor



Tampone rettale screening CRE



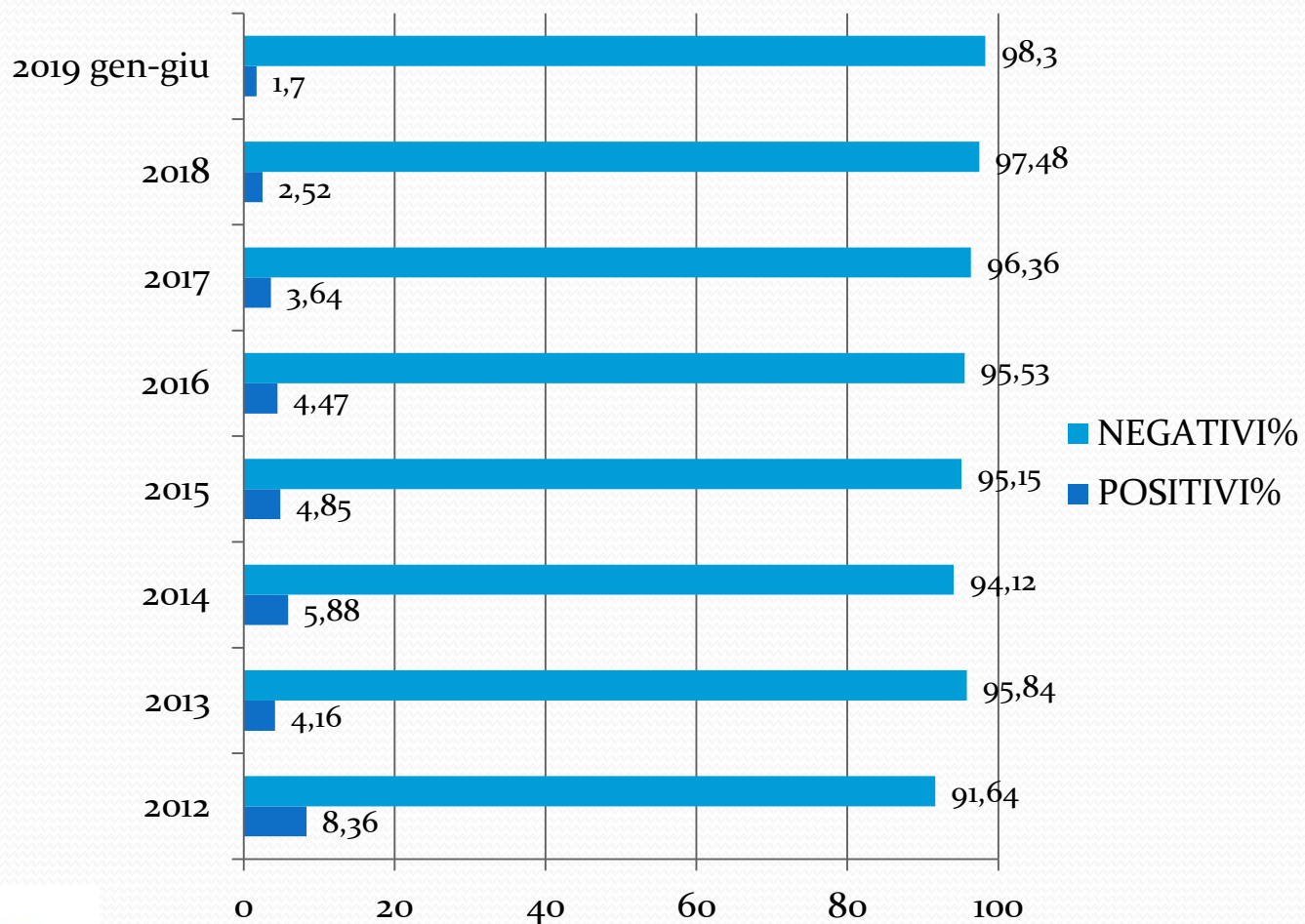
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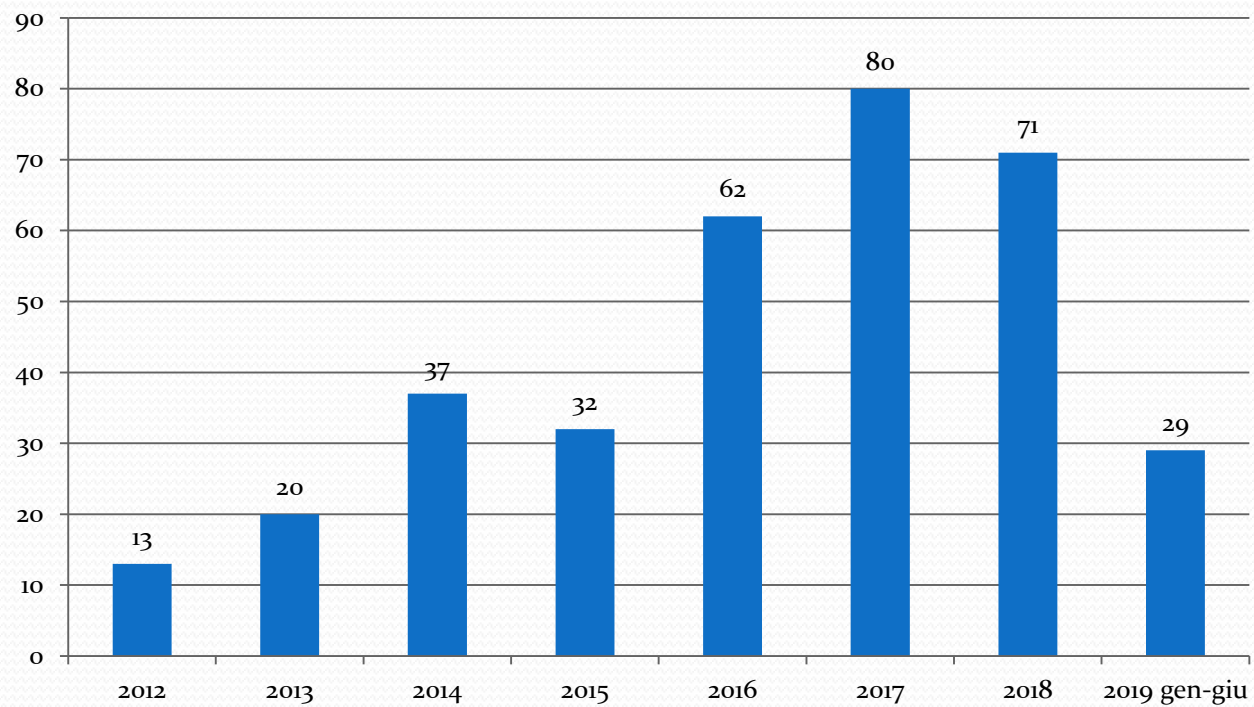


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PAZIENTI POSITIVI



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Colonizzazione intestinale da *Klebsiella pneumoniae* produttore di carbapenemasi e rischio di infezione da germi *multi-drug resistant* (MDR): studio retrospettivo.

Sala E.*, Mauri R., Cimetti S.°, Bonvini L.°, Busnelli M.°, Santoro D.

U.O. Malattie Infettive, U.O.S. Microbiologia*, Ufficio Epidemiologico Aziendale° – A.O. Sant'Anna - COMO

INTRODUZIONE La colonizzazione intestinale da *K. pneumoniae* produttore di carbapenemasi (KPC) è un problema emergente in ambito ospedaliero. In Europa, Grecia (>50%) e Italia (25-50%) presentano, in base ai dati EARSS 2011-2014, le maggiori percentuali di resistenza di *K. pneumoniae* ai carbapenemi. Presso l'Ospedale Sant'Anna di Como, il primo riscontro di KPC risale all'agosto 2010. Dall'agosto 2012 è in atto una sorveglianza attiva, con tampone rettale al momento del ricovero, per il rilievo dei pazienti colonizzati a livello intestinale con *Enterobacteriaceae* produttori di carbapenemasi. In questo studio ci proponiamo di definire se i soggetti portatori di KPC mostrano un aumentato rischio per infezioni e/o colonizzazioni sostenute da altri germi MDR.

METODI Lo studio caso-controllo è riferito al periodo 1 gennaio – 31 dicembre 2014, arruolando 34 pazienti risultati colonizzati allo screening e 34 pazienti di controllo negativi allo screening. I tamponi rettali sono stati eseguiti al momento del ricovero e la colonizzazione da KPC è stata sospettata in presenza di inibizione della azione di meropenem (alone di inibizione della crescita <30mm) su agar Mc Conkey. Le colonie sospette sono state identificate con il sistema Vitek 2 (bioMérieux) e con lo stesso sistema è stato eseguito l'antibiogramma secondo le regole EUCAST. La produzione di carbapenemasi è stata confermata con test fenotipici: test di Hodge modificato e Metallo-beta lattamasi/KPC test (Rosco Diagnostica).

RISULTATI 26 pazienti positivi allo screening per KPC hanno presentato una colonizzazione o infezione da germi MDR: *P. aeruginosa* (10 isolati), MRSA (4), *E. coli* ESBL (6), *C. difficile* (2), *P. mirabilis* ESBL (3), *A. baumannii* (3) e VRE (1). 8 pazienti positivi allo screening per KPC non presentavano infezioni o colonizzazioni da MDR, mentre 8 pazienti negativi allo screening per KPC mostravano colonizzazione o infezione da MDR: *P. aeruginosa* (1 isolato), MRSA (3), *E. coli* ESBL (3), *C. difficile* toxin positive (2), *P. mirabilis* ESBL (1), *K. pneumoniae* ESBL (1). 26 pazienti negativi allo screening per KPC risultano negativi anche per colonizzazione e infezioni da MDR.

CONCLUSIONI La colonizzazione intestinale da KPC è un fattore di rischio per la presenza di altri MDR: Odd Ratio=17 e $p=0,001$. In base ai dati raccolti nel nostro studio caso-controllo, c'è una correlazione altamente significativa tra colonizzazione intestinale da KPC e presenza di germi MDR. Questi risultati confermano l'utilità della sorveglianza attiva in ambiente ospedaliero, allo scopo di isolare i pazienti e prevenire la diffusione ospedaliera di KPC ed altri MDR.

Nosocomial Neonatal Outbreak of *Serratia marcescens* – Analysis of Pathogens by Pulsed Field Gel Electrophoresis and Polymerase Chain Reaction

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Outbreaks of *Serratia marcescens* in neonatal and pediatric intensive care units: Clinical aspects, risk factors and management

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Rapidly controlled outbreak of *Serratia marcescens* infection/colonisations in a neonatal intensive care unit, Pescara General Hospital, Pescara, Italy, April 2011

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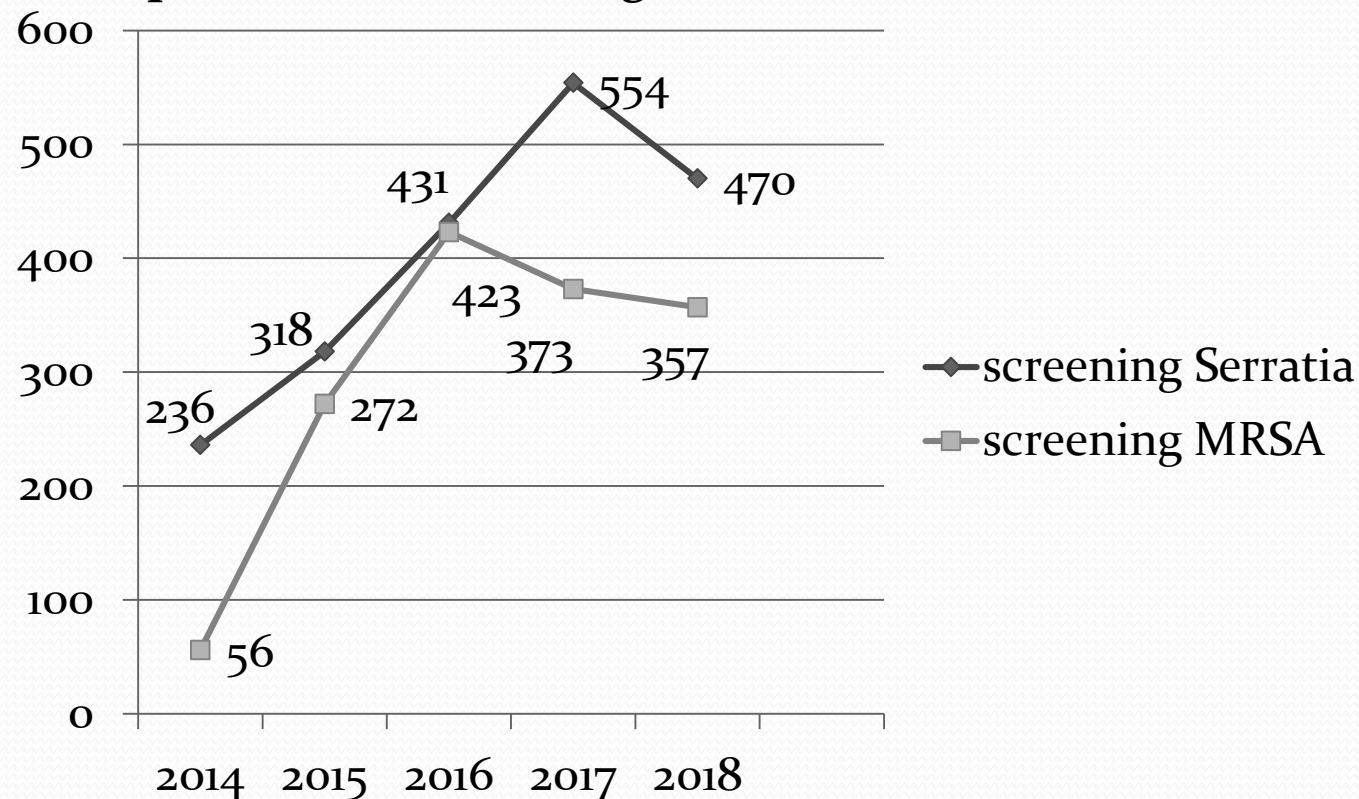
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TERAPIA INTENSIVA NEONATALE

Tampone faringeo e rettale screening Serratia

Tampone nasale screening MRSA



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